Carrier Screening for Thalassaemia and Hemoglobinopathies Beyond 2014

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Photographer : Joseph Benchapol

Scope of my talk

- Why do we need a new model for thalassemia carrier screening
- Pre-marital & reproductive screening
- Newborn screening
- Future plan

Prevalence of thalassaemia and hemoglobinopathy in South East Asian countries



Viprakasit V et al. Expert Opinion on Orphan Drug 2014

The scatter plots of MCV and MCH in individuals with β thalassemia with and without α thalassemia



Shaded boxes represent upper-lower range of values in β thalassemia trait with normal α genes ($\alpha\alpha/\alpha\alpha$) with the black lines denote the average (X) in each group. The black symbols represent individual cases with β thalassemia traits and α^{0} thalassemia while the white symbols signify β thalassemia traits with α^{+} thalassemia.

Viprakasit V et al. CCLM 2013

Current obstacle in prevention and control program for severe thalassemia in Thailand



Reported by Bureau of Health Promotion, Department of Health

The 13th Thailand National Thalassemia Conference 2007

Siriraj Hospital 's experience on couples at risk' decision regarding PND counseling



Vip Viprakasit & Chanin Limwongse, unpublished data 2012

Although the number of the participant couples was on the rise, detected fetus with severe thalassemia syndrome was **very low (<20%)**

Logical steps of prevention & control program for severe thalassemia

- Identification of disease carrier in a population
- Education on them on thalassemia/disease
- Dissuasion of marriage between
- to start the program? Alternative options or adoption
- nen and Wingenetic diagnosis
- P₁enatal diagnosis and counseling-Termination of pregnancy

What to concern when you start to screen for thalassemia carriers

- Screening program should be supported by public education and regulatory structures
- All medical personnel should empower individuals to make informed decision
- We must ensure that people are protected against discrimination as a result of their test

"Whom" should be screened...

- Cascade screening
- Antenatal screening
- Premarital screening
- School children

Cascade screening:

Nory offective in Cardinia Darkistan

Antenatal screening:

Must have PBD to offer

Premaritall screening:

- How to access them
- Is Discrimination against carrier females

- Yes (Haemoglobinopathies & Tay Sachs) Mitchelle JJ et al 1999 (Montreal Canada)
- Yes. Even 15 years after screening(Haemoglobinopathies)

Lena-Russo D et al 2002 (Marseille France)

Best not done

Frumkin & Ziatogora 2008 (Israel)

Our Future Paradigm Shift: Udom Bay Project A new model for Thal Screening

My "Framingham Hb Study" at Auwudom project Start in 2012









- DNA diagnosis for 13 common α-that mutations
 - DNA diagnosis for 8 common β-that mutations#
 - MLPA and direct DNA sequencing

vs. Normal

* = --^{SEA}, --^{THAI}, --^{FIL}, -- ^{MED}, - $\mathbf{\alpha}^{20.5}$, - $\mathbf{\alpha}^{3.7}$, - $\mathbf{\alpha}^{4.2}$ CD59, Hb Constant Spring, Hb Pakse, CD30, initiation codon, Hb Quong Sze.

=-28, CD8/9(+G), CD17(A-T), IVSI-I(G-T), IVSI-5, (G-C), CD26(HbE), CD41/42(-TCTT), CD71/72(A+), IVSII-654(G-T), 105bp, 619bp, 3.5kb, SEA HPFH, Filipino,

Hb Lepore, Asian Indian deletions inversion, Chinese, HPFH-6, Siriraj-thalassemia.



Genotypes

Newborn Screening: Because you touch the future everyday

If Untreated, Disorders

Can result in:

- Growth problems
- Developmental delays
- Behavioral/emotional problems
- Deafness or blindness
- Retardation
- Seizures
- Coma, sometimes leading to death



Newborn Screening (cord blood)

Can we diagnose thalassaemia at birth

Back to the "Future"



WHY Newborn Screening P

1.Newborn screening Program has been in existence but only for prevention and control of mental retardation.

2.The good period for Hb Bart's detection in α-thalassemia diagnosis.
> 1 year of age can be detected only by DNA analysis.

3.Detection of Hb E at birth.

Newborn screening for Thalassemia and hemoglobinophathies

Methodology :

High performance liquid chromatography (HPLC) Isoelectric focusing (IEF)

Advantage of IEF beyond HPLC method :

Can be used for both screening and confirmation. Applicable for newborn screening specimens (DBS). Efficiency of cost for screening. High resolution for Hb variants detection.

The Registry and Surveillance System for Hemoglobinopathies or RuSH,

- The National Heart, Lung, and Blood Institute (NHLBI)/NIH and the Division of Blood Disorders (DBD) at CDC develop a state-based monitoring system for SCD and thalassemia to identify and gather information on all people living with a hemoglobinopathy diagnosis of sickle cell diseases or thalassemia
- The participating states (California, Florida, Georgia, Michigan, North Carolina, Pennsylvania, and New York)
- during 2004-2008.

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Managing incidental findings and research

The Registry and Surveillance System for Hemoglobinopathies or RuSH,

- A pilot program that supported the development of the infrastructure and data collection methods for a state-based surveillance system for selected hemoglobinopathies.
- Total 31,144 individuals who had a hemoglobinopathy diagnosis during the study period were identified in California; 39,633 in Florida; 20,815 in Georgia; 12,680 in Michigan; 34,853 in New York, and 8,696 in North Carolina.
- State health department employees, healthcare providers, academic institutions, community organizations, patients, and families were all important contributors to the program, and they worked closely with each other throughout the entire process.



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Diagnostic applications of newborn screening for α -thalassaemias, haemoglobins E and H disorders using isoelectric focusing on dry blood spots

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Pronpanich, P et al. Ann Clin Biochem 2013

Schematic Diagram of Research Planning



1. To set up a standard range of each hemoglobin profile identified at newborn period for definitive diagnosis for thalassemia diseases and carriers.

2. To determine the sensitivity and specificity of IEF for thalassemia diseases and carriers identification in the newborns in Thailand.

Materials and methods

Subjects

350 dried blood specimens of newborn infants from newborn unit, Pediatrics, Phramongutklao hospital, Bangkok, Thailand.





IEF for Neonatal Screening using dry blood spots



Pronpanich, P et al. Ann Clin Biochem 2013



Pronpanich, P et al. Ann Clin Biochem 2013

Sensitivity and specificity of IEF for thalassemia diseases and carriers identification in the newborns in Thailand

Sensitivity: 100 %

Specificity: 100%

β/β					β ^Ε /β ^Ε					
•	a a /a a Hb Bart's Hb E	-a/aa	/a a	 -a 0	a a /a a	-a/a a	/a a	-a <i>1</i> -a	aajaa	/a a
<u>8.7</u>	2		(China)				90	0		0
•1.1		°				• 00 0		•	0	
• 0									Ø	

Sensitivity: 82.1 % Sensitivity: 92.9 %

Specificity: 7.1 % Specificity: 82.1 %

Pronpanich, P et al. Ann Clin Biochem 2013

The effect of α -globin gene deletion to Hb E levels

The effect of Hb E mutation to the amount of Hb Bart's

Cut off level for heterozygous α-thalassemia2 diagnosis

ROC curves for the diagnosis of heterozygous α -thalassemia-2, based on Hb Barts levels. (Area under curve 0.835).

Neonatal screening with IEF can diagnose Hb E, EE & β^0 thal/Hb E <u>+</u> α thalassemia

Table 4 Standard reference range for presumptive diagnosis of α -thalassaemia traits, Hb H and Hb E disorders using the concentrations of Hb Bart's and Hb E.

Hb Bart's					
Hb E(A2)	0-0.25	0.25-1.1	5.2-8.7	23.0-33.0	Interpretation
0-1.0	αα/αα, β^{A} / β^{A}	-α/αα, β^{A}/β^{A}	/αα, β ^A / β ^A	/-α, β ^A / β ^A	Genotypes
	Normal	α^+ thal trait	α ⁰ thal trait	Hb H dis.	Phenotypes
1.5–1.7		-a /aa, $\beta^{\rm E}$ / $\beta^{\rm A}$	/αα, $β^{E}/ β^{A}$ - α/-α, $β^{E}/ β^{A}$	/- α , β^{E}/β^{A}	Genotypes
		Hb E and α^+ thal trait	Hb E and α^0 thal trait	Hb E trait and Hb H dis	Phenotypes
2.0-4.5	αα/αα, β ^E / β ^A				Genotypes
	Hb E trait				Phenotypes
4.8-5.6	αα/αα, $\beta^{\rm E}$ / $\beta^{\rm E}$	-α/αα, β^{E}/β^{E}	/αα, β ^E / β ^E	/-α, β ^Ε / β ^Ε	Genotypes
	Homo E	Homo E and α^+ thal trait	Homo E and α^0 thal trait	Homo E and Hb H dis.**	Phenotypes

*Known as Hb AE Bart's disease.

**Known as Hb EF Bart's disease.

Hb Bart's ranges and associated α-globin gene status depending on hemoglobin plateforms

	Tanphaichitr VS, et al. Birth Defect (1988)		Sriroongrueng W, et al. Southeast Asian J Trop Med Public Health (1997)		M.J. Rugless, et al. Hemoglobin (2006)		Charoenkwan P, et al. Blood Cells, Molecules, and Diseases (2010)		Present Study	
	N	% Hb Bart's	N	% Hb Bart's	Ν	% Hb Bart's	N	% Hb Bart's	N	% Hb Bart's
Total	406		285		103		562		350	
Normal a-globin gene	326	0-4.9	238	0	30	0.5-5.6	399	0-0.1	306	0-1
Single <i>a</i> -globin gene defect	63	0-3.7	31	0-5.30	46	1.5-5.7	120	0-1	22	0-1.5
Two a-globin genes defect	16	4.1-9.8	14	2.29-9.00	27	6.1-11.9	34	3.2-12.9	19	5.2-8.7
Deletional Hb H disease	N	lot available	2	21.05-21.07	N	ot available	9	23.4-29.4	3	23-33
Hb Bart's Hydrops fetalis	1	97.5		Not available	Not available		Not available		Not available	
Hb Isolation Technique	Starch ;	gel EP	Starch	gel EP	HPLC		IEF		IEF	
Hb Measurement Technique	Hb Measurement Technique Cellulose acetate		Cellulose acetate		HPLC		Isoscan®		Isoscan®	
Molecular technique Southern blot hybridization		Southern blot hybridization		Southern blot hybridization, α-globin sequence analysis		Multiplex PCR		Multiplex PCR, PCR-RFLP		

Diagnosing heterozygous β- thalassemia in the newborns

Fig. 4. Distribution of HbA₁ values at birth (734 entries). The peak centered around 9% (horizontal stripes) corresponds to β -thalassamic heterozygous, the one at 20% (vertical stripes) to normal newborns.

Gianfranco C., et al. J Hematol. (1982)

Diagnosing heterozygous β- thalassemia in the newborns

Comparison of Hb A level of normal newborn and the newborns with heterozygous β-thalassemia

Stability of hemoglobin in dried blood spot studied by IEF

Neonatal screening with IEF can diagnose Hb E, EE & β^0 thal/Hb E <u>+</u> α thalassemia

Hb E mutation	α-globin gene	Charoenkwan P, et al. (2010)				Present Study			
		Ν	%Hb Bart' s	% Hb E	Ν	%Hb Bart' s	% Hb E		
	Normal	41	0-0.3	0.9-2.9	72	0-1	2.0-4.5		
e ^E /e	Single gene defect	19	0-0.13	0-0.5	5	0.3-1.0	1.5-1.7		
рγр	Two genes defect	2	4.2, 12.9	1.8, 0.5	5	5.9-6.9	1.5-1.7		
	Three genes defect	2	23.8, 29.4	1.0, 2.1		NA			
e ^E /e ^E	Normal		ΝΑ		3	0-0.6	5.1-5.6		
Р \ Р	Two genes defect		INA		1	6.8	4.8		
β ⁰ /β ^E	Normal	1 - 2.2 NA							

Summary

- There is limiting efficacy of our current P&C program in antenatal period
- Carrier detection program can be performed at other groups: reproductive & newborn screening
- NB screening provides a potential effective model for future Thai generation as it can be incorporated into our existing NB screening program
- Further study is underway to provide a comprehensive management scheme for NB-4-Thal Health Service

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CDC: Center for Disease Control & Prevention, USA

- Routine testing of all newborns for some of the hemoglobinopathies is performed by the state-based newborn screening (NBS) programs since 2006
- Although screening for sickle cell disease (SCD), one of the hemoglobinopathies, has been included as part of NBS in all 50 states, screening for α- and β-thalassemia, is currently performed in only a few states.
- In addition, many people at risk for a hemoglobinopathy who live in the United States were born either before NBS began in their state, or in a country without NBS.
- For these reasons, the actual number of people in the United States with hemoglobinopathies, and the associated public health impact, are unknown.

CDC: Center for Disease Control & Prevention, USA

There is no ongoing monitoring system for hemoglobinopathies. This lack of a monitoring system makes it difficult to:

- Identify people with these conditions,
- Monitor use of healthcare services and any resulting changes in health or quality of life, and
- Understand the impact of these conditions on the healthcare system.